

Product Sheet



Luciferase Daudi(CD20 KO) Cell Line

Catalog number: GM-C48708

Version 4.0.26.929

Specifications

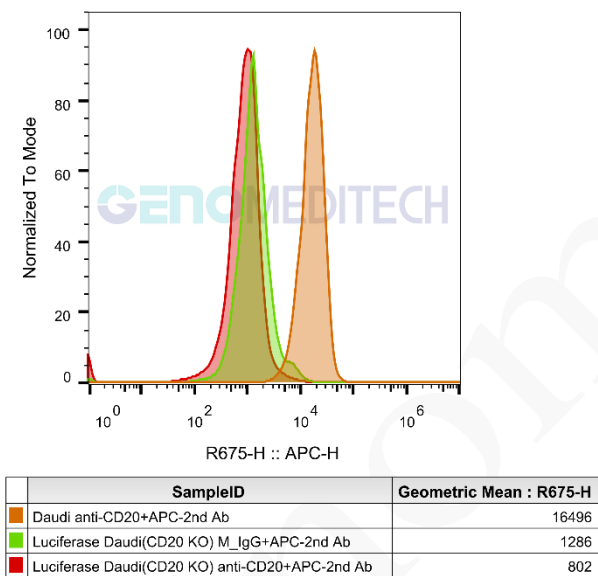
Description	Luciferase Daudi(CD20 KO) Cell Line is a clonal stable Daudi (Human CD20 KO) cell line that constitutively expresses the Luciferase gene, constructed using lentiviral technology.
Quantity	5E6 Cells per vial, 1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	Human CD20
Gene ID/Uniprot ID	/
Host Cell	Daudi
Recovery Medium	RPMI 1640+10% FBS+1% P.S
Growth medium	RPMI 1640+10% FBS+1% P.S+1.5 µg/mL Blasticidin+0.5 µg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Suspension
Growth Conditions	37°C, 5% CO ₂
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

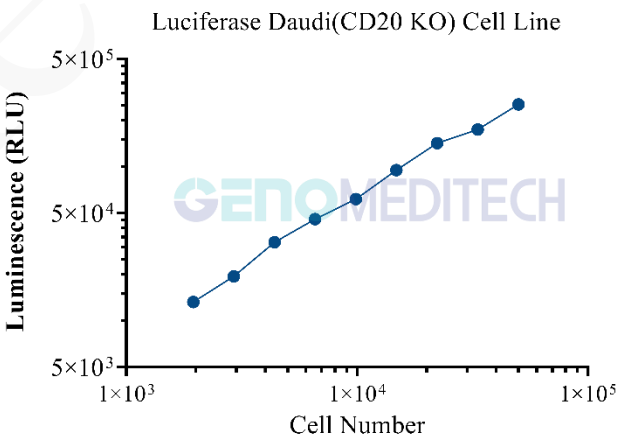
Reagent	Manufacturer/Catalogue No.
RPMI 1640	gibco/C11875500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ GM-040404
Puromycin	Genomeditech/ GM-040401
Anti-CD20 mIgG2a Antibody (ocrelizumab)	Genomeditech/GM-88845AB
GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit	Genomeditech/ GM-040513
Countess 3 Automated Cell Counter	ThermoFisher/A49891
NovoCyte Flow Cytometer	Agilent/NovoCyte 2060R

Data

Expression

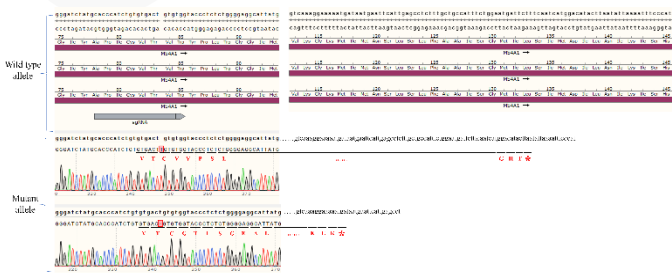


Luciferase Daudi(CD20 KO) Cell Line (Cat. GM-C48708) was determined by flow cytometry using Anti-CD20 mIgG2a Antibody (ocrelizumab) (Cat. GM-88845AB).



Correlation between the number of cells and bioluminescence values. Serial dilutions of Luciferase Daudi(CD20 KO) Cell Line (Cat. GM-C48708) (96-well format). The firefly luciferase activity was measured using the GMOne-Step 2.0 Luciferase Reporter Gene Assay Kit (Cat. [GM-040513](#)).

Genotype Verification



The Sanger sequencing of the Luciferase Daudi(CD20 KO) Cell Line showed successful knockout of CD20.

Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at 176 x g for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.
- Aliquot 1 mL into each vial.
- Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+1.5 µg/mL Blasticidin+0.5 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- After the first resuscitation, the cells can be subcultured after approximately 2–3 days. After 1 to 2 passages, the culture medium can be changed to a growth medium supplemented with antibiotics. If passage is not possible within 3 days, it is recommended to supplement with recovery medium as appropriate and place the flask in a horizontal position.
- When the cell density reaches 8E5 cells/mL, perform a 1:3 split, and continue subculturing every 3–4 days. Do not allow the cell density to exceed 1E6 cells/mL. It is recommended to use T25 flasks for passaging and culture.
- These cells are suspension cells, and it is recommended to use the "half-medium change" method to maintain optimal cell conditions during passaging.
- During passaging, you can directly add fresh growth medium to the culture flask, gently pipette to resuspend the cells, and then transfer the cell suspension to a new T-25 flask for continued culture.

Subcultivation Ratio: Maintain cultures at a cell concentration between 2.5E5 and 8E5 viable cells/mL.

Medium Renewal: Every 3 to 4 days

Notes

- It is recommended to increase the serum concentration to 20% during cell resuscitation.

- b) Fetal bovine serum (FBS) should be heat-inactivated at 56°C for 30 minutes, which can deactivate complement and some viruses without significantly affecting the activity of most growth factors and cytokines.

Related Products

CD20(MS4A1)	
H_CD20 KO RAMOS Cell Line	Luciferase Raji (CD20 KO) Cell Line
ADCC FcγRIIIa(158V) Jurkat Effector Cell Line	Cynomolgus_CD20 CHO-K1 Cell line
H_CD20 CHO-K1 Cell Line	H_CD20 HEK-293 Cell Line
Mouse_CD20 CHO-K1 Cell Line	
Anti-CD20 hIgG1 Reference Antibody (Ocrebio)	Anti-CD20 hIgG1 Reference Antibody(Ritubio)
Anti-CD3×CD20 hIgG1 Bispecific Antibody (Epcobio)	Anti-H_MS4A1(CD20) hIgG1 Antibody(Ocrelizumab)
CD19	
H_CD19 KO Raji Cell Line	Cynomolgus_CD19 CHO-K1 Cell Line
Cynomolgus_CD19 HEK-293 Cell Line	H_CD19 CHO-K1 Cell line
H_CD19 HEK-293 Cell Line	Mouse_CD19 CHO-K1 Cell Line
Anti-CD19 hIgG1 Reference Antibody (Loncbio)	Anti-CD19×CD3 hIgG1 Antibody[PIT-565(CD58 K34A)]
Anti-H_CD19 hIgG1/hIgG2 Antibody(Tafasitamab)	
CD3	
Cynomolgus_CD3D CD3E Reporter Jurkat(CD3DE KO) Cell Line	Jurkat CD3-BsAb Reporter Cell Line
Cynomolgus_CD3 HEK-293 Cell Line	Cynomolgus_CD3E(Membrane Bound ECD) CHO-K1 Cell Line
H_CD3 CHO-K1 Cell Line	H_CD3 HEK-293 Cell Line
H_CD3(TCR V2) CHO-K1 Cell Line	H_CD3(TCR V2) HEK-293 Cell Line
H_CD3(TCRαβ delta V) HEK-293 Cell Line	H_CD3D CD3E KO Jurkat Cell Line
H_CD3E KO Jurkat Cell Line	H_CD3E(Membrane Bound ECD) CHO-K1 Cell Line
Mouse_CD3 HEK-293 Cell Line	
Anti-CD19×CD3 hIgG1 Antibody[PIT-565(CD58 K34A)]	Anti-CD3 epsilon hIgG1 Antibody [OKT-3 (muromonab)]
Anti-CD3 hIgG1 Antibody(CH2527)	Anti-CD3×CD20 hIgG1 Bispecific Antibody (Epcobio)
Anti-CD3×FCRL5 hIgG1 Bispecific Antibody(cevostamab)	Anti-CD3E×BCMA hIgG4 Reference Antibody (Tecbio)
Anti-CD3E×DLL3 hIgG1 Bispecific Antibody(Tarlabio)	Anti-CD3E×MUC17 hIgG1 Bispecific Antibody(Vepsitbio)
Anti-mouse CD3ε mIgG2a Antibody(145-2C11)	
Human CD3E Protein; hFc Tag	

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