

Product Sheet



H_FOLH1(PSMA) KO LNCap Cell Line

Catalog number: GM-C45888

Version 4.0.26.923

Specifications

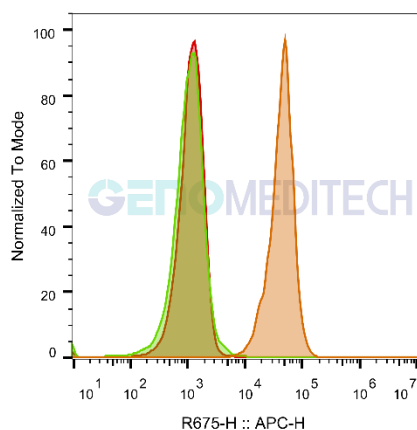
Description	H_FOLH1(PSMA) KO LNCap Cell Line is a clonal stable cell line derived from LNCap cells with a knockout of human FOLH1.
Quantity	3E6 Cells per vial, 1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Target	H_FOLH1(PSMA)
Gene ID/Uniprot ID	/
Host Cell	LNCap Cell Line
Recovery Medium	RPMI 1640+15% FBS(Gibco)+1% P.S
Growth medium	RPMI 1640+15% FBS(Gibco)+1% P.S
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO ₂
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
RPMI 1640	gibco/C11875500BT
Fetal Bovine Serum	Gibco/10099-141C
Pen/Strep	Thermo/15140-122
Anti-FOLH1(PSMA) hIgG1 Antibody(Rosopatamab)	Genomeditech/ GM-27353AB
NovoCyte Flow Cytometer	Agilent/NovoCyte 2060R
Countess 3 Automated Cell Counter	ThermoFisher/A49891

Data

Expression



	SampleID	Geometric Mean : R675-H
	LNCap Anti-FOLH1+APC 2nd Ab	42852
	H_FOLH1(PSMA) KO LNCap H_IgG+APC 2nd Ab	1037
	H_FOLH1(PSMA) KO LNCap Anti-FOLH1+APC 2nd Ab	1144

The Sanger sequencing of the H_FOLH1(PSMA) KO LNCap Cell Line showed successful knockout of FOLH1.

Genotype Verification



H_FOLH1(PSMA) KO LNCap Cell Line is a clonal stable cell line derived from LNCap cells with a knockout of human FOLH1.

Cell Recovery

Recovery Medium: RPMI 1640+15% FBS(Gibco)+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately 176 x g for 5 minutes. Discard supernatant.
- d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into an appropriate culture flask and initially place the flask in an upright position after thawing.
- e) Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at 176 x g for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 3E6 cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: RPMI 1640+15% FBS(Gibco)+1% P.S

Antibiotics are not required during cell culture. The composition of the growth medium is identical to that of the recovery medium.

- a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:2 every 4-5 days. Ensure that the density does not exceed 90%, as overcrowding can lead to reduced viability due to compression.
- b) Remove and discard culture medium.
- c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes at 37°C).
- e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:2 is recommended

Medium Renewal: Every 4 to 5 days

Notes

- Cells immediately after thawing typically contain a certain proportion of dead cells, which is a normal phenomenon. With appropriate adjustment and recovery, cell viability generally improves markedly. Once the culture stabilizes, the proportion of dead cells decreases after subsequent passages, and the cell growth rate becomes more consistent.
- Conventional TC-treated culture vessels provide suboptimal substrate for cell adhesion. To improve attachment efficiency, the surface should be pre-conditioned with poly-L-lysine solution before seeding, or Corning CellBIND surface culture flasks (Cat. No. 3289) should be used as an alternative.
- FBS should be heat-inactivated at 56 °C for 30 minutes in a water bath. This process inactivates complement and some viruses, while generally having minimal impact on the activity of most growth factors and cytokines.

Related Products

FOLH1(PSMA)	
Cynomolgus_FOLH1(PSMA) CHO-K1 Cell Line	H_FOLH1(PSMA) CHO-K1 Cell Line
H_FOLH1(PSMA) HEK-293 Cell Line	H_FOLH1(PSMA) PC-3 Cell Line
H_FOLH1(PSMA) RM-1 Cell Line	
Anti-FOLH1(PSMA) hIgG1 Antibody(Rosopatamab)	Anti-FOLH1(PSMA) hIgG1 Reference Antibody (Rosobio)
Cynomolgus FOLH1(PSMA) Protein; His Tag	
STEAP1	
H_STEAP1 KO LNCaP Cell Line	Cynomolgus_STEAP1 CHO-K1 Cell Line
H_STEAP1 CHO-K1 Cell Line	H_STEAP1 HEK-293 Cell Line
H_STEAP1 RM-1 Cell Line(Low Expression)	Mouse_STEAP1 CHO-K1 Cell Line
Rat_STEAP1 CHO-K1 Cell Line	
Anti-H_STEAP1 hIgG1 Antibody(Vandortuzumab)	Anti-STEAP1 hIgG1 Reference Antibody (Vandbio)
ADC Related Product	
Anti-DXD Mouse IgG1 Antibody (23E21C5)	Anti-DXD Mouse IgG1 Antibody (4A5A12)
Anti-Dxd Mouse IgG2a Antibody (17D6A4)	Anti-Eribulin Mouse IgG2a Antibody (10F8G4)
Anti-MMAE Mouse IgG1 Antibody (11C10E3)	Anti-MMAE Mouse IgG2a Antibody (17A1K11)
Anti-MMAE Mouse IgG2a Antibody (8F6A3)	Anti-SN38 Mouse IgG1 Antibody(59H11C7)
Mouse anti Human IgG1-DXD(Dar8)	Mouse anti Human IgG1-MMAE(Dar4)
Human IgG1 Isotype-DXD (Dar8)	Human IgG1 Isotype-Eribulin (Dar4)
Human IgG1 Isotype-MMAE (Dar4)	
Recombinant DT3C Protein	
AR(NR3C4)	
H_AR Reporter 293 Cell Line	H_AR HEK-293 Cell Line
KLK2 KLK3	
Membrane bound Cynomolgus_KLK2(AA25-270)-Flag CHO-K1 Cell Line	Membrane bound H_KLK2(AA19-261) 22RV1 Cell Line
Membrane bound H_KLK2(AA19-261) CHO-K1 Cell Line	Membrane bound H_KLK2(AA19-261) HEK-293 Cell Line

Membrane bound H_KLK2(AA19-261) LnCap Cell Line	Membrane bound H_KLK2(AA19-261) Luciferase LnCap Cell Line
Membrane bound H_KLK2(AA25-261) CHO-K1 Cell Line	Membrane bound H_KLK2(AA25-261) CT26 Cell Line
Membrane bound H_KLK2(AA25-261) HEK-293 Cell Line	Membrane bound H_KLK2(AA25-261) MC38 Cell Line (Low Expression)
Anti-KLK2 hIgG1 Antibody(Hu11B6)	
ACP3	
Flag-H_ACP3 HCT116 Cell Line	Flag-H_ACP3 HT-1080 Cell Line
H_ACP3 22Rv1 Cell Line	H_ACP3 CHO-K1 Cell Line
H_ACP3 HCT116 Cell Line	H_ACP3 HEK-293 Cell Line
H_ACP3 HT-1080 Cell Line	H_ACP3 PC-3 Cell Line

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